

## **EFFECT OF CADMIUM ON MORPHOMETRIC AND BIOCHEMICAL PLANT PARAMETERS OF *TRIGONELLA FOENUM - GRAECUM L.***

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### **Introduction**

In an agriculture-based country like India, livestock production is of utmost vitality and plays a key role in providing employment especially in Indian agriculture and rural economy. Livestock adds 9% to National GDP and 25% to agricultural GDP respectively and contribution from livestock is about 15-20% to the farmer's household income (Hegde, 2006). Nutrition plays a major role by driving the functional efficacy, and efficiency of the livestock production system. The fodder from common cereal crops are rich in energy and the leguminous crops are rich in proteins. The green fodder crops are known to be the best source of nutrients as compared to concentrates and hence useful in bringing down the cost of feeding and reduce the need for the purchase of feeds from the market (Roy et al., 1993).

Heavy metal contamination of soil is a far more serious problem than air or water pollution because heavy metals are usually tightly bound by the organic components in the surface layers of the soil. Consequently, the soil is an important geochemical sink which accumulates heavy metals quickly and usually depletes them very slowly by leaching into groundwater aquifers or bioaccumulating in plants (Infotox, 2000). Heavy metals can also be very quickly translocated through the environment by the erosion of the soil particles to which they may adsorb or bound and redeposited elsewhere. Industrial processing and intensive agricultural practices, resulting in the contamination of forage, feed, and water, are sources of Cd exposure for farmed ruminants. From here, they migrate into the food chain by direct or indirect usage of respective crops. Although some heavy metals like Cu, Fe, Mn, Zn are required for the growth of plants in

trace amounts but prove fatal if present beyond their maximum permissible limits (Freitas et al., 2010). Various heavy metals along with Cd, viz., arsenic, copper, cobalt, lead, manganese, mercury, nickel, and zinc are reported to cause genotoxicity upon reaching the living systems (Chandra et al., 2005; Bertin et al., 2006).

Cd accumulates over time, primarily in the kidney and liver (Langlands et al., 1988). Cd can cross various biological membranes and, once intracellular, to bind to ligands with exceptional affinity. Cd is a known human carcinogen (Filipic et al., 2006). Exposure to low Cd concentrations can also adversely affect bovine reproduction (Kreis et al., 1993). Cd contamination in food crops and their effects on human health have not been extensively reported. An improved understanding of the interaction of Cd with other elements may provide the key to understanding the effects of Cd on health. Therefore, in the present investigation, the effects of heavy metal Cadmium (Cd) on fodder plant *Trigonella foenum-graecum* from Rajasthan state was studied.

## **Material and methods-**

Certified seeds of *Trigonella foenum-graecum* were obtained from Durgapura Agriculture Research Station, Jaipur. The seeds were stored in sterilized glass stoppered bottles.

### **1. Morphometric evaluation of plant parameters under Cd stress-**

Uniform, healthy and viable seeds were taken for the experiment. The seeds were surface sterilized with 0.01% mercuric chloride solution for three minutes and thoroughly washed with distilled water three times each for 5 minutes and then dried with a paper towel. Dry seeds were placed in 90-mm-diameter Petri dishes on a layer of Whatman No. 1 filter paper with a thin uniform pad of sterilized cotton placed beneath, and then moistened with 6 different cadmium sulfate concentrations (i.e., 10, 50, 100, 200, 500 and 1000 ppm), and seeds also grown in distilled water as a control. Seeds were kept at room temperature ( $25^{\circ}\text{C} \pm 1^{\circ}\text{C}$ ) under normal light for germination. Each treatment includes 3 Petri dishes as replicates which contain 10 healthy and homogenous seeds (10 Seeds/ Petri dish)

arranged in a concentric ring and equidistant from each other on the filter paper. 30 seeds of *Trigonella foenumgraecum* were used. The emergence of radical (not less than 2 mm) was considered an indication of successful germination. The number of germinated seeds was counted daily until there was no further seed germination. The total number of seeds germinated was recorded. The length of plumule and radical was recorded on the 10th after germination. For radical and plumule length and fresh and dried weight average of 10 seedlings was taken. The data thus obtained were used to calculate germination percentage, the kinetics of plumule emergence, seedling length, fresh and dry weight, growth index, and seed vigor. Following formulae were used to calculate the parameters:

$$(a) \text{ Seed germination} = \frac{\text{No. of Seed germination}}{\text{Total no. of seed sowed}}$$

**(b) Shoot and Root Length (cm)-**

The length of root and shoot were separately measured of 10 days old seedlings with the help of a ruler (in cm).

**(c) Fresh and dry weight (gm)-**

After proper washing, excess water was removed and then the fresh weight of seedling was measured, and for dry weight the seedling was dried in an oven at 80°C for 24 hours and then weighed.

**2. Biochemical evaluation of plant parameters under Cd stress–**

For biochemical evaluation, seeds were sown in experimental pots and hydroponic systems in the Department of Botany, Rameshwari Devi Girls College, Bharatpur (Raj.). The various attributes considered for the present work were studied at a different time interval after germination. The following attributes were analyzed for fenugreek crop:

**(a) Protein estimation-**

The leaf proteins of control and treated plants extracted by Lowry et al. 1951 method. Alkaline  $\text{CuSO}_4$  catalyzes the oxidation of aromatic amino acids with subsequent reduction of sodium-potassium molybdate tungstate of Folin's reagent giving a purple color complex the intensity of the color is directly

proportion to the concentration of the aromatic amino acid in the given sample solution. For extraction, grind 500mg of 10 days old plant sample with pestle and mortar in 5-10 ml of the phosphate buffer. Centrifuge the homogenate at 8000 rpm for 20 min. Collect the supernatant and repeat the extraction 4-5 times. Combine the supernatants and make the volume to 50 ml with phosphate buffer. Take 0.1 ml of the above extract and add 0.1 ml of 20% TCA. Keep it for half an hour and centrifuge at 8000 rpm for 20 min. Wash the pellet with acetone twice and again centrifuge it. Discard the supernatant. Dissolve the pellet in 5 ml of 0.1 N NaOH and mix well till it gets dissolved. For estimation, a suitable aliquot (1 ml) of the above solution is taken and added to it 4.5 ml of freshly prepared alkaline copper sulfate reagent. Mix properly and after 10 min add 0.5 ml of Folin's reagent. Mix the content instantaneously. Allow the color to develop for 30 min. Record the absorbance at 660 nm after setting the instrument with reagent blank which contains 1 ml of 0.1 N NaOH instead of the sample aliquot. In another set of tubes take suitable aliquots of BSA solution (in a range of 10-100 µg) make the total volume to 1 ml with 0.1 N NaOH and develop the color as described above. Draw a standard curve of absorbance at 660 nm versus µg of BSA. From the standard graph the amount of protein in the given unknown solution is calculated and expressed as mg gm<sup>-1</sup> fw of leaves.

**(b) Total soluble sugar-**

Total soluble sugar was estimated by using anthrone reagent (Dubois et al., 1951). 25mg of the sample was crushed in 10 ml of 80% ethanol and then centrifuged for 10 minutes at 4000 rpm. Take the supernatant as an extract. Add 0.1 ml of the extract to 4 ml of the anthrone solution and heated for 10 minutes in boiling water and was allowed to cool at room temperature. The intensity of blue-green color was measured in UV-VIS spectrophotometer at 625 nm against a reagent blank (anthrone reagent was responsible for color development). The sugar content was estimated from a standard curve prepared with a known concentration of glucose.

**(c) Estimation of Phenols –**

The deproteinized test materials (200mg each) were macerated with 10 mL of 80% ethanol for 2 hours, and left overnight at room temperature. The mixtures were centrifuged, and the supernatants were collected separately and

maintained up to 40 mL by adding 80% ethanol. Total phenol content in each sample was estimated by the spectrophotometer method of Bray and Thorpe, 1954. It includes the preparation of a regression curve of a standard phenol (Tannic acid). A stock solution of tannic acid was prepared by mixing 40 mg of standard phenol in 1 mL of 80% ethanol. Eight concentrations ranging from 0.1 to 0.8 mL were prepared in the test tube and volume was raised to 1 mL by addition of 80% ethanol. To each test tube, 1 mL of Folin-Ciocalteu reagent (commercially available reagent was diluted by distilled water in 1:2 ratio just before use) and 2 mL of 20% sodium carbonate solution was added and the mixture was shaken thoroughly. The samples were placed in boiling water for 1 min and cooled under running water. These reaction mixtures were diluted to 25 mL by adding distilled water and optical density was read at 750 nm against a blank. The optical density of each sample was plotted against the respective concentration of total phenols to compute the regression curve. The concentrations in the test samples were calculated by referring the respective optical density of the test sample against a standard curve of tannic acid (or standard phenol).

#### **(d) Lipid Estimation-**

The test sample was dried, powdered and 100 mg was macerated with 10 mL distilled water, transferred to a conical flask containing 30 mL of chloroform and methanol (2/1: v/v; Jayaram, 1981). The mixture was thoroughly mixed and left overnight at room temperature in dark for complete extraction. Later, 20 mL of chloroform mixed with 2 mL of water was added and centrifuged. Two layers were separated, the lower layer of chloroform, which contained all the lipids, was carefully collected in the pre-weighed glass vials and the colored aqueous layer of methanol which contained all the water-soluble substances and thick interface layer were discarded in each test sample. The chloroform layers were dried in vacuum and weighed. Each treatment was repeated thrice and their mean values were calculated.

## **RESULTS AND DISCUSSION-**

The accumulation of heavy metals in agricultural soils is of increasing concern due to the food safety issues and potential health risks as well

asits detrimental effects on soil ecosystems (Qishlaqi and Moore, 2007). The heavy metal concentration in the soil solution plays an important role in controlling metal bioavailability to plants. Presently, due to constraints in the availability of freshwater for irrigation, wastewater is being used for irrigation of agricultural fields resulting in toxic metal contamination. Heavy metals are either leach into ground or surface water and enter into the growing food crops (Janos et al., 2010). From here, they migrate into the food chain by direct or indirect usage of respective crops. Although some heavy metals like Cu, Fe, Mn, Zn are required for the growth of plants in trace amounts but prove fatal if present beyond their maximum permissible limits (Freitas et al., 2010). Various heavy metals along with the Cd, viz., arsenic, copper, cobalt, lead, manganese, mercury, nickel, and zinc are reported to cause genotoxicity upon reaching the living systems (Chandra et al., 2005; Bertin et al., 2006).

Therefore, in the current investigation, the effect of cadmium stress on morphometric and biochemical plant parameters of fodder plant *Trigonella foenum-graecum* was tested. For that, seeds of *Trigonella foenum-graecum* sown in 3 treatment groups of pots of triplicates of three experiments each were observed for every undertaken experiment.

## **1. Morphometric observation of plant parameters under Cadmium stress**

### **(a) Seed germination percentage-**

Seed germination is a vital developmental event in plants and considered an important growth stage that is frequently subjected to high mortality rates. The percentage of germination has been used as an indicator of heavy metal toxicity in plants. The effect of metals on the development and reproduction of plants can be firstly quantified by determining the germination characteristics of seeds (Munzuroglu and Geckil, 2002). The effect of heavy metals on seed germination depends on their ability to penetrate the seed coat and disturb various physiological processes involved in germination (Khattab, 2004; Faheed, 2005; Wang and Zhou, 2005).

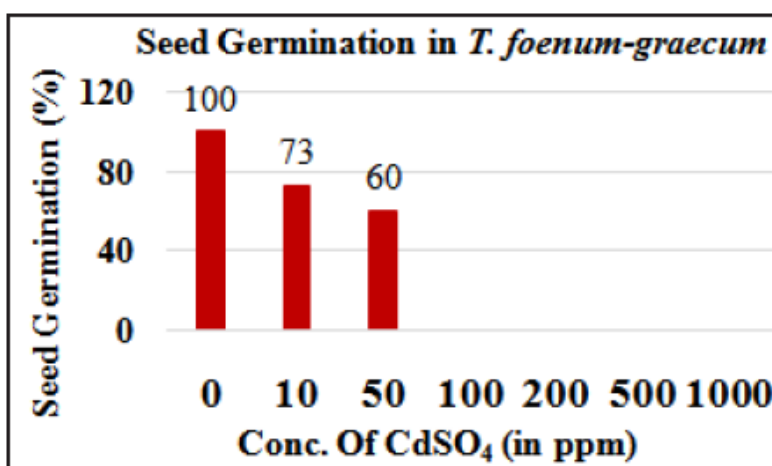
The seeds of *Trigonella foenum-graecum* were screened for cadmium salt tolerance deriving germination percentage. Results indicated that Cadmium stress given in the form of Cadmium sulfate had a significant inhibitory effect on

germination percentage. By increasing concentrations of cadmium sulfate, the germination percentage gradually decreased concerning the control in the groups treated with increasing concentrations of cadmium sulfate from 10, 50, 100, 200, 500, and 1000 ppm. Treatment on the seeds of *Trigonella foenum-graecum* exhibited only 73% germination at 10 ppm, 60% at 50 ppm, and null at 100 ppm or higher treatment. However, 100% germination under control condition when cadmium sulfate was absent (0 ppm). The effect of cadmium stress on seed germination of *Trigonella foenum-graecum* is shown in Table-1 and Figure-1(a), 1(b)

The inhibition percentage of germination in *Trigonella* was high in heavy metals stress. In high-level treatments, germination percentages were detrimentally affected, implying that a higher concentration of cadmium was not conducive to seed germination. This may be attributed to depression of oxygen uptake and physiological disturbance in the mobilization of reserved food materials (Mondal et al., 2013).  $CdCl_2$  and  $HgCl_2$  (0.05-50 mM) have reduced seed germination and the shoot development with an increase of metal concentrations and exposure time (Alhelal, 1995).

**Table-1: Effect of Heavy Metal Cd on Seed Germination(%) in *Trigonella foenum-graecum* (Values are means of three replicates each)**

Concentration of $CdSO_4$ (in ppm)	0	10	50	100	200	500	1000
Seed Germination (%)	100	73	60	-	-	-	-



**Figure-1 (a): Effect of Heavy Metal Cd on Seed Germination (%) in *Trigonella foenum-graecum***



**Figure-1 (b): *Trigonella foenum-graecum* seeds under Cd stress of varied concentration**

### (b) Shoot Length

In the present study, the results recorded significant reductions in shoot length with all the treated groups from 10, 50, 100, 200, 500, and 1000 ppm concentrations of cadmium sulfate salt as compared with the control in *Trigonella foenum-graecum*. Shoot length was decreased from 4 cm in control to 2.4 cm and 1.1 cm at 10 and 50 ppm respectively. No growth was observed at higher concentrations (100, 200, 500, and 1000 ppm) of cadmium sulfate. The effect of cadmium stress on shoot length of *Trigonella foenum-graecum* has been shown in Table-2 and Figure-2.

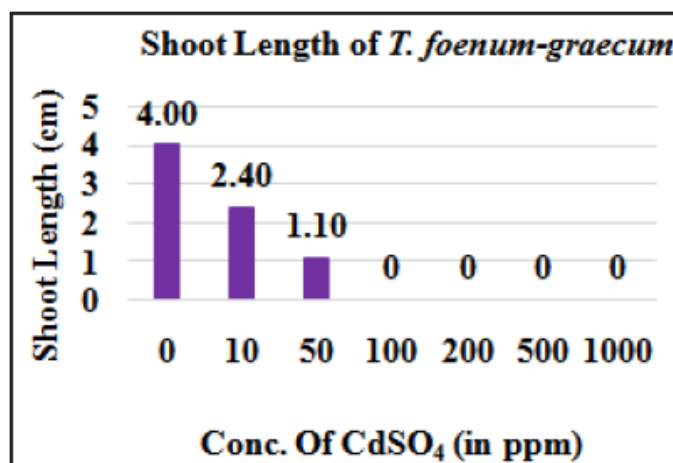
The reasons for the inhibition effect of heavy metal to plant growth were probably due to a series of physical and chemical reactions between excess heavy metal and soil components which changes soil properties, thus affecting soil fertility levels (Cieslinski et al., 1996; Chang and Wu, 2005). However, the level of significance varied in different varieties, cadmium sulfate levels, and at different



durations. Mukherjee and Dalal, 2014 demonstrated that root and shoot length was drastically changed under heavy metal stress. After the accumulation of heavy metals, the length of root and shoot were decreased. There was a decrease in root and shoot length in *Trigonella foenum-graecum* under lead and cadmium stress. Pb and Cd treated *Trigonella foenum-graecum* showed a decreased level of tolerance indices which also dose-dependent and Cd shows the maximum effect for the decreased root length. Perveen et al., 2011 also showed that Cd treated *Trigonella foenum-graecum* was exhibit blackening of the root system (Godbold and Kettner, 1991). Ahmad et al., 2005 reported that there was a significant decrease in root and shoot length in lead and cadmium treated *Trigonella*.

**Table-2: Effect of Heavy Metal Cd on Shoot Length (cm) of Seedlings in *Trigonella foenum-graecum* (Values are means of three replicates each)**

Concentration of CdSO <sub>4</sub> (in ppm)	0	10	50	100	200	500	1000
Shoot Length (cm)	4	2.4	1.1	-	-	-	-



**Figure-2: Effect of Heavy Metal Cd on Shoot Length (cm) of Seedlings in *Trigonella foenum-graecum***

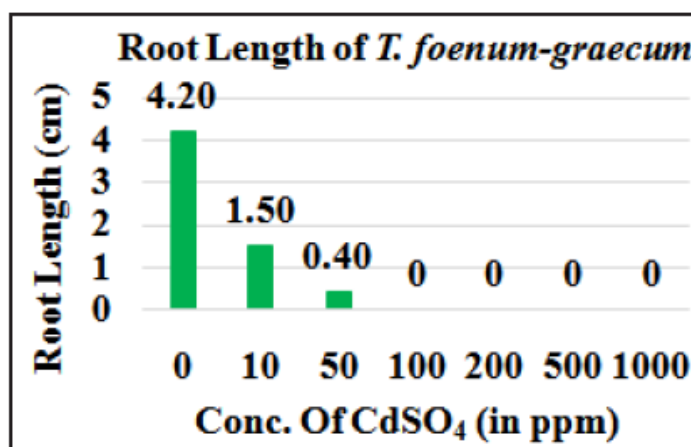
**(c) Root Length**

In the present study, similar to shootlength, root length has also been adversely affected undertreated concentrations of cadmium sulfate. Under nonstressed conditions, when cadmium sulfate was absent (0ppm), root length in *T. foenum-graecum* was 4.2 cm. In treated groups, it was noticed that the root length was decreased at 10 ppm and 50 ppm to 1.5 cm and 0.4 cm, and further, there was no growth was observed. The effects of cadmium stress on root length of *Trigonella foenum-graecum* have been shown in Table-3 and Figure-3.

In the present study, similar to shoot length, root length has also been adversely affected under various concentrations of cadmium sulfate. Zayneb et al., 2015 reported that cadmium affected various plant growth parameters. Its accumulation was markedly lower in shoots as compared to roots, reducing root biomass by almost 50%. Chugh and Sawhney, 1995 also investigated the deleterious effect of cadmium on fenugreek  $\alpha$ - and  $\beta$ -amylase, which were quite sensitive to metal: both were affected equally by 35% and 46% in the presence of 0.25 mM and 0.50 mM cadmium, respectively.

**Table-3: Effect of Heavy Metal Cd on Root Length (cm) of Seedlings in *Trigonella foenum-graecum* (Values are means of three replicates each)**

Concentration of CdSO <sub>4</sub> (in ppm)	0	10	50	100	200	500	1000
Root Length (cm)	4.2	1.5	0.4	-	-	-	-



**Figure-3: Effect of Heavy Metal Cd on Root Length (cm) of Seedlings in *Trigonella foenum-graecum***

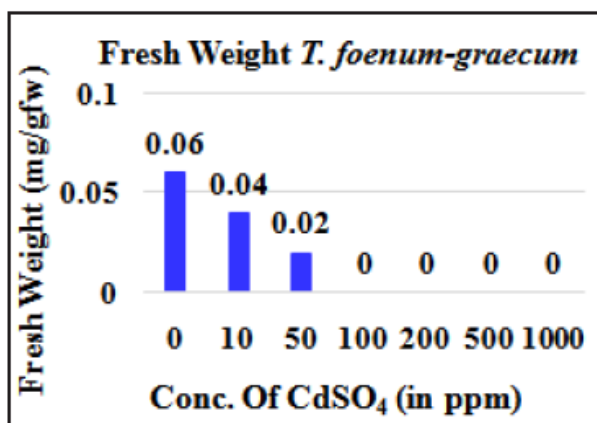
**(d) Fresh weight of seedlings-**

The fresh weight of untreated seedlings in *Trigonella foenum-graecum* was 0.06g. A decrease in weight was observed with 0.04 and 0.02 g at 10 and 50 ppm and further no growth was observed at higher concentrations. The effects of cadmium stress on the fresh weight of *Trigonella foenum-graecum* are shown in Table-4 and Figure-4.

The fresh weight of seedlings in *Trigonella foenum-graecum* was significantly affected by increasing Cd dosage. Pirselova et al., 2016 reported that each of the tested doses of Cd resulted in a decrease of root fresh weight by 31.70 and 28.68%. Biyani et al., 2019 also described that Cd stress caused a significant decrease in shoot fresh and dry weight. The fresh and dry biomass accumulation of Cd-treated plants was significantly lower than in plants that were adequately supplemented with Fe (+Fe/+Cd). The greatest reduction was observed in “Fe/+Cd plants after 10 days of treatment, whereas this effect was reversed after the addition of Fe to the plants (+Fe/+Cd), even in the presence of Cd treatment.

**Table-4: Effect of Heavy Metal Cd on Fresh Weight (g) of Seedlings in *Trigonella foenum-graecum* (Values are means of three replicates each)**

Concentration of CdSO <sub>4</sub> (in ppm)	0	10	50	100	200	500	1000
Fresh White (g)	0.06	0.04	0.02	-	-	-	-



**Figure-4: Effect of Heavy Metal Cd on Fresh Weight (g) of Seedlings in *Trigonella foenum-graecum***

## 2. Biochemical observation of plant parameters under Cadmium stress

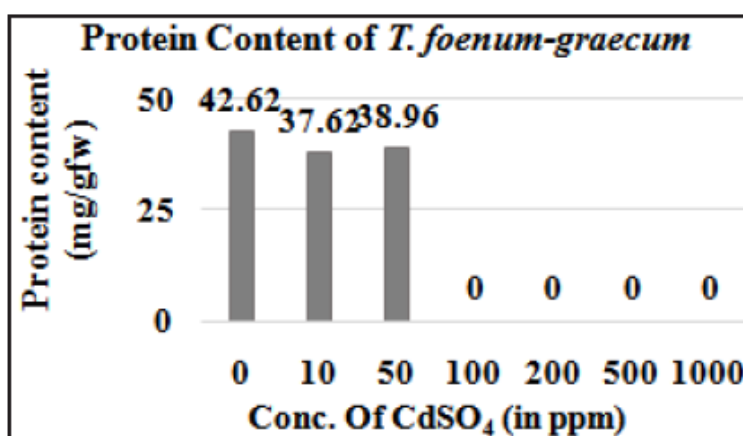
### (a) Protein content-

There was a slight decrease in the protein content of *Trigonella foenum-graecum* from control 42.62mg/g at 10 ppm level (37.62 mg/g) and the slight increase at 50 ppm level (38.96 mg/g) was obtained compared to control. The effects of cadmium stress on the protein content of *Trigonella foenum-graecum* is shown in Table-5 and Figure-5

Abiotic stress may inhibit the synthesis of some proteins and promote others (Ericson and Alfinito, 1984). In different crop plant heavy metal toxicity also reduced the level of proteins (Tamas et al., 1997). Various types of grains developed on the cadmium treated soils also show a lower level of protein content (Salgare and Achareke, 1992). Cd stress could trigger the production of free ROS and result in oxidative damage in plants.

**Table-5: Effect of Heavy Metal Cd on Protein content (mg/g) in the Seedlings of *Trigonella foenum-graecum* (Values are means of three replicates each)**

Concentration of CdSO <sub>4</sub> (in ppm)	0	10	50	100	200	500	1000
Fresh White (g)	42.62	37.62	38.96	-	-	-	-



**Figure-5: Effect of Heavy Metal Cd on Protein content (mg/g) in the Seedlings of *Trigonella foenum-graecum***

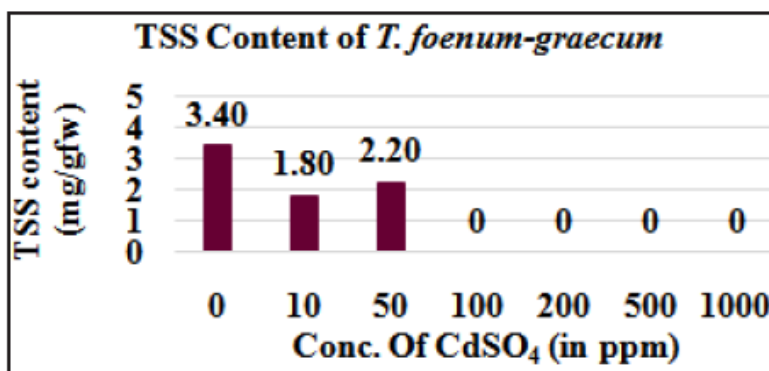
**(b) Total Soluble Sugar content**

The soluble carbohydrate content in *Trigonella foenum-graecum* was higher at non treated conditions with 3.4 mg/g. Initially the decrease in the TSS content was obtained in *Trigonella foenum-graecum* with 1.8 mg/g and 2.2 mg/g at 10 and 50 ppm level and further no content was exhibited at higher concentrations (100, 200, 500 and 1000 ppm concentrations) of Cd stress. The effects of Cd stress on the total soluble sugar content of *Trigonella foenum-graecum* is shown in Table-6 and Figure-6.

Sugar metabolism has adversely affected the plants grown under stressed conditions (Jha and Dubey, 2004). Saleh and Garni, 2006, reported that more than 5 mg/kg concentration of Cd inhibited the total carbohydrate content. Ci et al., 2009, also demonstrated that the total soluble sugar concentration decreased in four wheat (*Triticum aestivum* L.) lines differing in cadmium (Cd) tolerance were subjected to 50 µM CdCl<sub>2</sub> from the three-leaf stage for 24 days.

**Table-6: Effect of Heavy Metal Cd on TSS content (mg/g fresh weight) in the Seedlings of *Trigonella foenum-graecum* (Values are means of three replicates each)**

Concentration of CdSO <sub>4</sub> (in ppm)	0	10	50	100	200	500	1000
TSS Content	3.4	1.8	2.2	-	-	-	-



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**Figure-6: Effect of Heavy Metal Cd on TSS content (mg/ g fresh weight) in the Seedlings of Trigonella foenumgraecum**

**(c) Phenol content-**

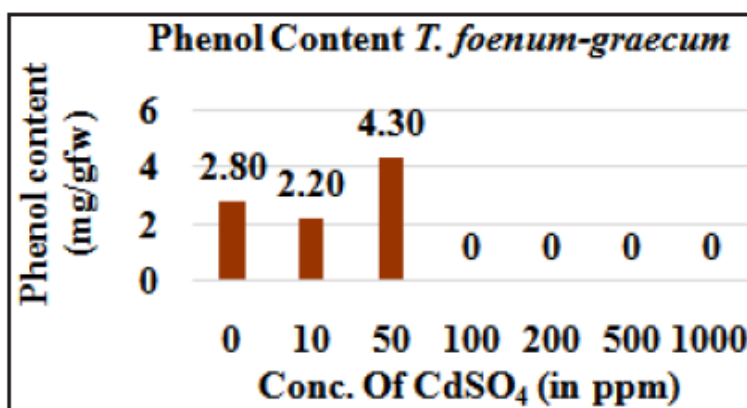
The phenol content at the control conditions of *Trigonella foenum-graecum* was 2.8 mg/g, it reduces to 2.2 mg/g at 10 ppm level, and then it was observed to increase at 50 ppm upto 4.3 mg/g which is greater than control. However, no phenol content was observed at higher Cd stress concentrations. The effects of Cd stress on phenol content of *Trigonella foenum-graecum* is shown in Table-7 and Figure-7.

Patel et al., 2013 results agree with the findings of Hamid et al., 2010 who found that the phenolic content of plants was decreasing with increasing levels of heavy metal. These reports justify the initial decline in the phenol content. Korat et al., 2019 also studied the total phenol content from seedlings (10, 20, and 30 days after germination) of different fenugreek genotypes that were found statistically significant for a different stage. Among the genotypes, the mean total phenol content varied from 0.424 to 0.570 mg g<sup>-1</sup>. In the case of 10 to 30 days after germination, there was an increase in the total phenol content in a leaf of all fenugreek genotypes. At 30 days, among the 10 genotypes the total phenol was remained significantly higher in JFG-80 (0.759 mg g<sup>-1</sup>) and the lowest was found in GM-2 (0.527 mg g<sup>-1</sup>). Mukherjee and Dalal, 2014, reported that lead and cadmium stressed *Trigonella* shows a marked increase level of total phenol in a dose-dependent manner and a higher concentration of cadmium shows maximum level. It was also revealed gradual accumulation of proline and total phenols in seedling leaves. Phenols in plants may perform useful effects through free radicals scavenging (Chun et al., 2003). This also reflects an increase in antioxidant activity and the reason why phenol content was increased at higher Cd levels.

**Table-7: Effect of Heavy Metal Cd on Phenol content (mg/g fresh weight) in the Seedlings of *Trigonella foenum-graecum* (Values are means of three replicates each)**

Concentration of CdSO <sub>4</sub>	0	10	50	100	200	500	1000
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(in ppm)							
Phenol content (mg/gfw)	2.8	2.2	4.3	-	-	-	-



**Figure-7: Effect of Heavy Metal Cd on Phenol content (mg/g fresh weight) in the Seedlings of Trigonella foenum-graecum**

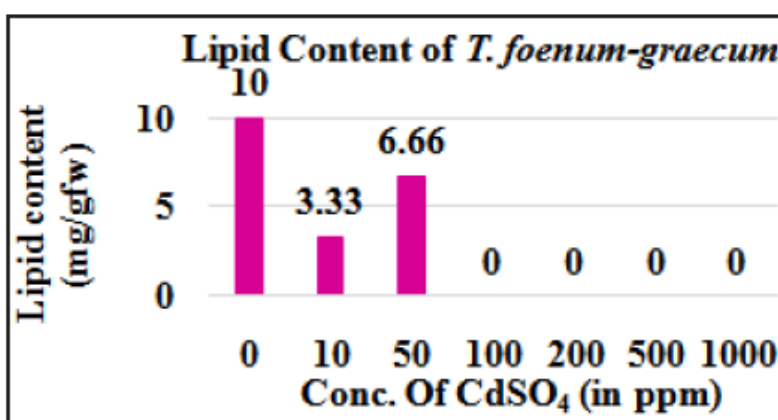
**(d) Lipid content-**

Lipid peroxidation (MDA) is a biochemical marker for free radical-mediated injury. Under the oxidative stress MDA is the final product of lipid peroxidation and the most prominent indicator of oxidative stress in various stressed plants (Sima et al., 2012). As per the results obtained in the present study, a decrease in lipid content was observed in *Trigonella foenum-graecum* with increasing Cd concentration. Lipid content of the control group was 10 mg/g which was reduced to 3.33 mg/g at 10 ppm but at 50 ppm it was found to be 6.66 mg/g and further no lipid content was determined at higher concentrations of Cd stress. The effects of Cd stress on the lipid content of *Trigonella foenum-graecum* have been shown in Table-8 and Figure-8.

In the study conducted by Zhan et al., 2017, the level of lipid peroxidation and cell death were higher in Cd-treated seedlings compared with those of the controls. Similarly, in another study as well, *Trigonella foenum-graecum* in response to cadmium stress showed significantly higher doses of MDA than the control and it was greatly affected by the highest doses of Cd. All the concentrations of Cd treatment effect MDA production.

**Table-8: Effect of Heavy Metal Cd on Lipid content (mg/g fresh weight) in the Seedlings of *Trigonella foenumgraecum* (values are means of three replicates each)**

Concentration of CdSO <sub>4</sub> (in ppm)	0	10	50	100	200	500	1000
Lipid (mg/gfw)	10	3.33	6.66	-	-	-	-



**Figure-8: Effect of Heavy Metal Cd on Lipid content (mg/g fresh weight) in the Seedlings of *Trigonella foenum-graecum***

## REFERENCES

1. Ahmad, S.H., Reshi, Z., Ahmad, J. and Iqbal, M. (2005):Morpho-anatomical responses of *Trigonella foenumgraecum*Linn. To induced cadmium and lead stress.J. of plant biology. Vol. 48(1), pp. 64-84.
2. Alhelal, A. A. (1995): Effect of cadmium and mercury on seed germination and early seedling growth of Rice and Alfalfa. J. Univ. Kuwait. Sci. Vol. 22, pp. 76-83.
3. Arnon DI. (1949). Copper enzymes in isolated chloroplasts, polyphenoxidase in *Beta vulgaris*. PlantPhysiol; 24:1-15.



4. Bertin G and Averbeck D. 2006. Cadmium: cellulareffects, modifications of biomolecules, modulation ofDNA repair and genotoxic consequences Biochimie,88, 1549-1559.
5. Biyani, K. Tripathi, D. K. Lee, J. H. Muneer S. 2019.Dynamic role of iron supply in amelioration of cadmiumstress by modulating antioxidative pathways and peroxidaseenzymes in mungbean. AoB Plants,Vol 11(2).
8. Chun OK, Kim DO, Lee CY. Superoxide radical scavengingactivity of the major polyphenols in fresh plums.Journal Agriculture and Food Chemistry. 2003;51:8067-8072.
9. Freitas M, Gomes A, Porto G and Fernandes E. 2010.Nickel induces oxidative burst, NF-kappa B activationand interleukin-8 production in human neutrophils,Journal Biology Inorganic Chemistry. 15, 1275-1283.
10. Hegde NG. "Positive Attitude for Good Health and Happiness".Nature Cure Ashram, Urulikanchan, Pune(2006).
11. Jha, A. B. and Dubey, R. S. (2004): Carbohydratemetabolism in growing ice seedlings under arsenictoxicity. J. Plant Physiol. Vol. 161, pp. 867-872.
12. Khattab, H. (2004) Metabolic and oxidative responsesassociated with exposure of Eruca sativa (Rocket)plants to different levels of selenium. International Journalof Agriculture & Biology 6: 1101-1106.
13. Kreis,I.A. , M. deDoes, J.A. Hoekstra, C. de LezenneCoulander, P.W. Peters, G.H. Wentink. Effects of cadmiumon reproduction, an epizootological study.Teratology, 48 (1993), pp. 189-196
14. Mondal, N. K. Das, C. Roy, S. Datta J. K. and BanerjeeA. (2013). Effect Of Varying Cadmium Stress OnChickpea (Cicer Arietinum L) Seedlings: An UltrastructuralStudy. Annals of Environmental Science, Vol 7,59-70.
15. National Research Council Mineral Tolerance of DomesticAnimals. National Academy of SciencesPress, Washington DC, USA (1980)