
Comparative Phytochemical profiling of various extracts, from different parts of *Sesuviumportulacastrum* using GCMS, FTIR and ICP AES.

Snehal B. Gagare¹,

Pratima S Jadhav²

Department of Biochemistry The Institute of Science, Mumbai, Maharashtra.

Abstract:

Sesuviumportulacastrum (Aizoaceae family) is mangrove associate known for its antimicrobial, antifungal activity. The present study focuses on comparative analysis of phytochemicals present in *Sesuviumportulacastrum* (Aizoaceae family) using GCMS, ICP AES and IR spectroscopy. The work also highlights the different phytochemicals present in various parts like stem and leaves of the plants from Aizoaceae family. The IR spectroscopy elucidates different functional groups present in the phytochemicals from different parts of plant. The GCMS has identified 15, 15, 14 phytochemicals respectively from *Sesuviumportulacastrum* methyl acetate leaf, petroleum ether stem and chloroform whole plant extracts. This study also is noteworthy as it uses ICP AES techniques in analyzing different elements present in *Sesuviumportulacastrum*. This work emphasizes the importance of sophisticated analytical instrumentation, in phytochemical characterization and the technique can also reveal the difference in phytochemicals present in different parts of the same plant.

Keywords: *Sesuviumportulacastrum*, GCMS, ICP AES, IR spectroscopy.

Introduction:

Plants are used by human beings for food, fodder and medicine since ancient times. Medicinal plants form a large group of economically vital plants which provide the basic raw materials for native pharmaceuticals.¹ The plants that flourish in stressful coastal environment in particular mangroves, are rich in synthesis of secondary metabolites to surmount the stress.² Mangrove and mangrove associates contain biologically active antiviral, antibacterial and antifungal compounds. They provide a rich source of steroids, triterpenes, saponins, flavonoids, alkaloids and tannins.^{3,4,5} In traditional medicines, mangrove have been used against human, animal and plant pathogen, but detailed investigations regarding the bioactive components are inadequate.⁶

Sesuviumportulacastrum (L.) L. (seapurslane) is one of the fast growing, herbaceous, perennial, dichotomous, halophyte belonging to family Aizoaceae. In India, it grows at the eastern and western coastal sides as a mangrove associate.⁷ The plant is used on the Senegal coast as a haemostatic and a decoction of it is considered to be the best known antidote for stings of venomous fish. Leaves have acidulous flavour of sorrel as well as antiscorbutic⁸. The essential oil extracted from the leaves of *Sesuvium*, revealed notable antibacterial activity against both gram-positive and Gram-negative bacteria and displayed significant antifungal

and antioxidant activity ⁹. The plant is known to contain a polysaccharide, which showed positive activity against HIV¹⁰. The plant has revealed its importance in effective removal of heavy metals such as cadmium, lead and arsenic from contaminated sites¹¹. *Sesuviumportulacastrum*, molecular phylogenetic studies using 18S rRNA gene sequence has revealed, its closely related to *Perkesiaaculata* (Cactaceae) ¹².

Knowledge of the chemical constituents of plant is essential, for the discovery of therapeutic drugs as well as for finding out new sources of such economic materials as tannins, oils, gums, precursors for the synthesis of complex chemical substances ^{13,14}. Phytochemical which possess many economical and physiological roles are widely distributed as plant constituents.

Sesuviumportulacastrum has plethora of secondary metabolites with specific pharmacological prospects and hence there is need to investigate the phytochemicals present in various plant extracts. The bioactivity of phytochemicals can vary significantly depending on, plant parts, tissue type and at times with the growth conditions, solvent used in extraction. It is very difficult to standardize any formulation if the exact composition of chemical constituents is not known. There is a need to study the various chemicals present in different parts of plant to evaluate its potential.

With the advancement in analytical technology, details about metabolites present in plants can be identified with help of instruments like GCMS and IR spectroscopy. The principle of IR spectroscopy is based on the fact that various functional groups in a chemical structure gives rise to characteristic bands both in terms of intensity and position (frequency)¹⁵. The present study deals with chemical identification of *Sesuviumportulacastrum* using GCMS, ICP AES and IR spectroscopy. This study gives a comprehensive result about the application of various chemical based analytical methods used for identification of various metabolites from different parts like whole plant, stem and leaves of *Sesuviumportulacastrum*.

2. Materials and Methods

2.1 Plant materials :*Sesuviumportulacastrum* was collected from Kelve beach, Thane District Maharashtra identified and authenticated.

2.2 Chemicals and Reagents: All the chemicals used were of analytical grade and were purchased from Hi Media and Merck.

2.3 Plant extract preparation: 100 g of dry leaf, stem and whole plant of *Sesuviumportulacastrum* were extracted with ethyl acetate, chloroform and petroleum ether by using soxhlet apparatus. The extraction was filtered and kept for 24 hours to evaporate. Analytical techniques like GCMS, FT-IR and ICP AES were used for further phytochemical analysis.

2.4 GCMS analysis: The GCMS analysis was performed on Thermo Scientific TSQ 8000 Gas Chromatograph - Mass Spectrometer. The MS part consists of Triple Quadrupole, is paired with the TRACE 1300 GC along with Auto-sampler for automated sample handling. It is equipped with EI Ion Source programmable to 350 °C. The Mass Range is 2.1100 amu. Gas Chromatograph Consists of Split/Splitless Injectors and multi-mode (including on-column) Programmed Temperature Vaporizing (PTV), column Temperature 400°C.

2.5 IR Spectroscopy analysis: The IR Spectroscopy was performed on Perkin elmer Inc. Spectrum RX FT-IR spectrophotometer. It has an autosampler with fibre optic interfaces and range of microscope. Spectral resolution is better than 0.8 cm^{-1} It is equipped with dynascan interferometer.

2.6 ICP AES analysis: The ICP AES analysis was performed on ARCOS, Simultaneous ICP Spectrometer. It has an RF generator with a maximum of 1.6 KW, Radial plasma carries out the ionization and peristaltic pump delivers sample into analytical nebulizer. Spectrometer has a wavelength range of 130nm to 770 nm. Molecules from sample break into respective atoms and recombine with plasma giving its characteristic wavelength detected by CCD device

Result and Discussion:

The phytochemicals in *Sesuviumportulacastrum* were subjected to GCMS, IR, ICP AES spectroscopy. Identification of components by GCMS was based on direct comparison of the retention times and mass spectral data with those for standard compounds from the NIST library. The NIST library is a database which contains exhaustive information about various chemical compounds. The GCMS has identified 15, 15, 14 phytochemicals respectively from *Sesuviumportulacastrum* methyl acetate leaf, petroleum ether stem and chloroform whole plant extracts.

GCMS chromatogram of ethyl acetate leaf extract : The phytochemicals present in the ethyl acetate leaf extract are described in table no.1 and Figure no 1 displays the GCMS chromatogram. Tricaproin is obtained with maximum area percentage of 33.09% in ethylacetate leaf extract.

GCMS chromatogram of Petroleum ether stem extract: The phytochemicals present in the Petroleum ether stem extract are described in table no.2 and Figure no 2 displays the GCMS chromatogram. Cannabinoltrifluoroacetate is obtained with highest area percentage of 9.78 and Phenol is obtained with highest area percentage of 10.28%.

GCMS chromatogram of whole plant chloroform extract: The phytochemicals present in the Petroleum ether stem extract are described in table no.3 and Figure no 3 displays the GCMS chromatogram. Phenol, 2,4 bis phenyl ethyl is obtained at highest area percentage of 19.24%.

FTIR analysis of *Sesuviumportulacastrum* leaf: The IR spectrum is shown in figure no 4. The *Sesuviumportulacastrum* leaf extract yielded maximum peak level 3356 cm^{-1} and minimum peak 779 cm^{-1} . The leaf extract yielded flavones functional group at 1637 cm^{-1} . FT-IR studies confirm the presence of functional groups in leaf SeSL extract listed in table no 4.

FTIR analysis of *Sesuviumportulacastrum* whole plant: The IR spectrum is shown in figure no 5. The *Sesuviumportulacastrum* whole plant extract yielded maximum peak level 3600 cm^{-1} and minimum peak 783 cm^{-1} . The leaf extract yielded flavones functional group at 1641 cm^{-1} . FT-IR studies confirm the presence of functional groups in whole plant S1 extract listed in table no 5.

FTIR analysis of *Sesuviumportulacastrum* stem: The IR spectrum is shown in figure no 6. The *Sesuviumportulacastrum* stem extract yielded maximum peak level 3356 cm^{-1} and minimum peak 898 cm^{-1} . FT-IR studies confirm the presence of functional groups in stem SeSS extract listed in table no 6.

ICP AES analysis of *Sesuviumportulacastrum* :Elements present in the given sample are Cd,Cl,Co,Cr,Cu,Fe,K,La,Li,Mg,Mn,Na,Ni,P,,Pb,S,Si,Sr,Ti,V,Yb,Y,Zn,Ag, Al, B, Ba,Br, Ca.

Discussion:

Sesuviumportulacastrum has many different phytochemicals and their content varies significantly depending on the plant part and solvents used for the extraction purpose. In the present study leaf, stem and whole plant of *Sesuviumportulacastrum* was subjected to phytochemical analysis using GCMS , ICP AES, IR spectroscopy.

In *Sesuviumportulacastrum* leaf, Stem and whole plant extracts compounds like Cyclopentasiloxane, Dibutyl phthalate are commonly found in all the plant parts. There are even certain phytochemicals like Tricaproin, Stigmasterol, Sitosterol, Cannabinol trifluoroacetate which are confined to specific tissues only.

Our results are in accordance with the previous report on GCMS analysis of volatile oil from *Sesuviumportulacastrum* by Mohamed Yacoob Syed Ali which showed the presence of Hexadecanoic acid with highest peak percentage of 10.2 .¹⁶

The petroleum ether stem extracts of *Sesuviumportulacastrum* also contain Stigmasterol which was also reported in the previous preliminary studies performed by Amad All Azzawi et al¹⁷. This study gives a clear-cut particulars of the phytochemicals present in Whole plant, stem and leaves of *Sesuviumportulacastrum*. Many of the functional present in *Sesuviumportulacastrum* leaf, stem and whole plant extract are similar like aliphatic group ,alcohol group but the leaf extract contains flavones. So that compound may be phenolics. This identification is possible using IR spectroscopy as it give rise to characteristic bands both in terms of intensity and position (frequency).^{15,18,19}

The emission spectroscopy uses the inductively coupled plasma to produce excited atoms and ions that emit electromagnetic radiation at wavelengths characteristic of a particular element. The intensity of this emission is indicative of the concentration of the element within the sample.

The results obtained in the above studies are noteworthy as there are many reports on preliminary phytochemical analysis of *Sesuviumportulacastrum*^{9,16,17}, but there are no reports on use of IR spectroscopy and ICP AES in identification of phytochemicals.

Conclusion:

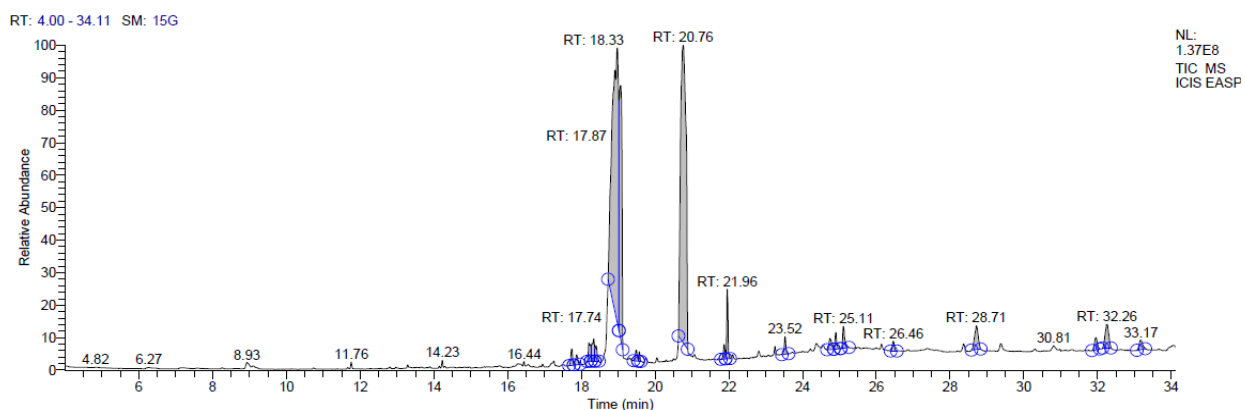
Phytochemical characterization of plant extract is essential, as it helps us to determine the exact composition of metabolites / chemicals in different plant .This kind of research also helps in correlating the chemical with their biological and physiological roles. The study also highlights use of sophisticated instruments like GCMS FTIR and ICP AES in phytochemical research to have immense knowledge of the plethora of chemicals present in different parts of the same plant.

Acknowledgment:

The authors are thankful to SAIF Chandigarh and IIT Powai for providing GCMS , ICP AES facility respectively.

Table no 1 : Retention time, Area%, Molecular formula and Major peaks of chemicals detected by GCMS of *Sesuviumportulacastrum*, ethyl acetate leaf extract (easp).

Peak no.	Rt	Area %	Molecular formula	Compound
1	8.22	1.55	C ₁₀ H ₃₀ O ₅ Si ₅	Cyclopentasiloxane
2	10.72	2.92	C ₁₄ H ₄₄ O ₆ Si ₇	Heptasiloxane,1,1,3,3,5,5,7,7,9,9,11,11,13,13tetradecamethyl
3	13.95	3.85	C ₁₆ H ₅₀ O ₇ Si ₈	Octasiloxane,1,1,3,3,5,5,7,7,9,9,11,11,13,13,15,15hexadecamethyl
4	14.95	4.52	C ₁₆ H ₄₈ O ₈ Si ₈	Cyclooctasiloxane, hexadecamethyl
5	16.67	1.02	C ₁₆ H ₄₈ O ₆ Si ₇	Heptasiloxane, hexadecamethyl
6	18.17	9.15	C ₁₆ H ₂₂ O ₄	Dibutyl phthalate
7	19.37	16.17	C ₁₉ H ₃₄ O ₂	9,12Octadecadienoicacid (Z,Z), methyl ester
8	19.97	5.85	C ₂₁ H ₃₈ O ₂	nPropyl 9,12octadecadienoate
9	20.76	33.9	C ₂₁ H ₃₈ O ₆	Tricaproin
10	23.21	3.66	C ₂₄ H ₃₈ O ₄	Diisooctyl phthalate
11	24.25	0.53	C ₂₀ H ₆₀ O ₁₀ Si ₁₀	Cyclodecasiloxane, eicosamethyl
12	26.66	1.70	C ₂₈ H ₃₉ ClO ₉	9Desoxy9xchloroingol 3,7,8,12tetraacetate
13	28.50	1.88	C ₂₃ H ₄₈	Heptadecane, 9hexyl28
14	30.17	1.67	C ₂₆ H ₅₄	Octadecane, 3ethyl5(2ethylbutyl)
15	33.07	2.56	C ₃₀ H ₅₀ O ₆	Olean12ene3,15,16,21,22,28hexol

Figure no 1: GCMS chromatogram of *Sesuviumportulacastrum*ethyl acetate leaf (EASP)**Figure no 2: GCMS chromatogram of *Sesuviumportulacastrum*petroleum ether stem(PEA4)**

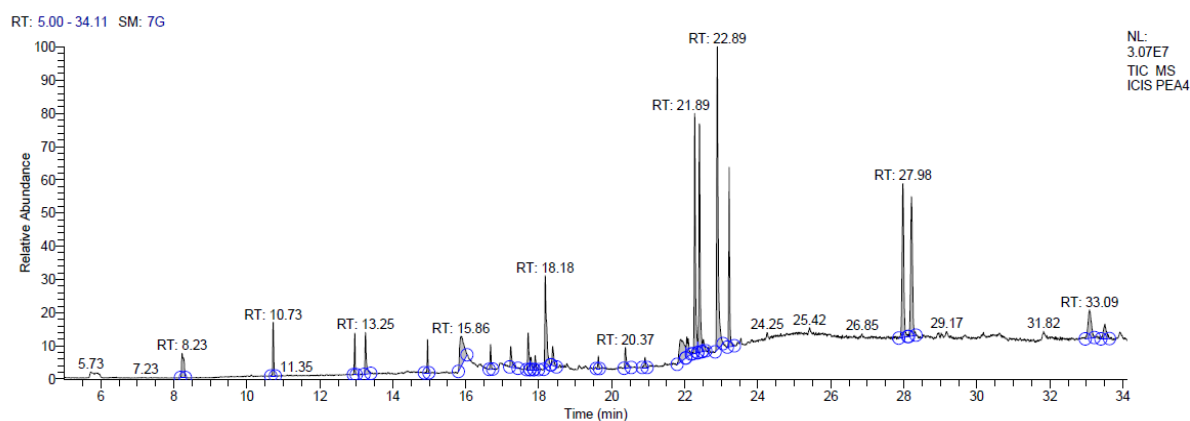


Figure no 3: GCMS chromatogram of *Sesuviumportulacastrum chloroform wholeplant (CLEA6)*

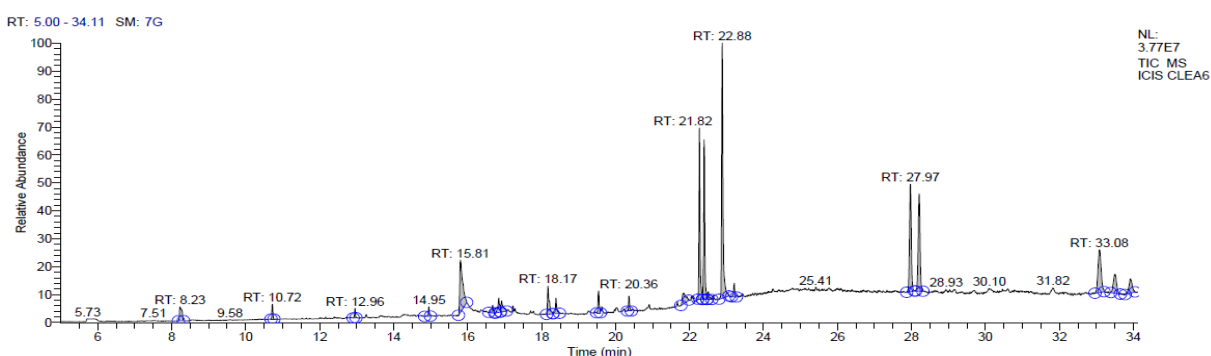


Table no2: Retention time, Area%, Molecular formula and Major peaks of chemicals detected by GCMS of *Sesuviumportulacastrum* Petroleum ether stem extract (pea4).

Peak no.	Rt	Area%	Molecular formula	Compound
1	8.23	2.03	C10H30O5Si5	Cyclopentasiloxane, decamethyl
2	10.73	1.87	C14H44O6Si7	Cyclohexasiloxane, dodecamethyl
3	13.25	2.05	C14H22O	Phenol, 2,4 bis(1,1 dimethylethyl)
4	15.86	4.38	C15H18	Azulene, 1,4dimethyl7(1methylethyl)
5	17.71	1.78	C16H22O4	Dibutyl phthalate
6	18.18	5.91	C16H22O4	Dibutyl phthalate
7	20.37	1.14	C16H34O2	Hexadecanoic acid, ethyl ester
8	21.89	3.89	C17H37N7O3	Deoxyspergualin
9	22.27	10.28	C14H22O	Phenol, 2,4 bis(1,1 dimethylethyl)
10	22.40	9.96	C14H22O	Phenol, 2,4 bis(1,1 dimethylethyl)
11	22.27	10.28	C22H22O	Phenol, 2,4bis(1phenylethyl)
12	23.22	6.48	C24H38O4	Pthalicacidi(2 propyl pentyl ester)
13	27.98	9.78	C23H25F3O3	Cannabinol, trifluoroacetate
14	28.21	9.44	C23H25F3O3	Cannabinol, trifluoroacetate
15	33.50	1.63	C29H48O	Stigmasterol

Table no3: Retention time, Area%, Molecular formula and Major peaks of chemicals detected by GCMS of *Sesuviumportulacastrum* whole plant chloroform extract (CLEA4).

Peak no.	Rt	Area%	Molecular formula	Compound
1	8.23	1.09	C10H30O5Si5	Cyclopentasiloxane, decamethyl
2	14.95	4.08	C14H42O5Si6	Hexasiloxane, tetradecamethyl
3	15.81	8.46	C14H14O	Phenol, 2(1phenylethyl)
4	16.84	1.43	C15H18	Azulene, 1,4dimethyl7(1methylethyl)
5	18.17	2.65	C16H22O4	Dibutyl phthalate
6	18.39	1.21	C18H36O2	Hexadecanoic acid, ethyl ester
7	19.54	1.40	C20H40O	Phytol
8	20.36	0.92	C22H44O2	Eicosyl acetate
9	21.82	2.12	C22H43NO	13Docosenamide,
10	22.27	10.72	C22H22O	Phenol, 2,4bis(1phenylethyl)
11	22.88	19.16	C22H22O	Phenol, 2,4bis(1phenylethyl)
12	27.97	10.74	C23H25F3O3	Cannabinol, trifluoroacetate
13	33.08	7.41	C29H50O	çSitosterol
14	33.91	2.53	C30H50O	áAmyrin

Table no 4: FTIR Result for *Sesuviumportulacastrum* leaf (SeSL)

Wavelength in cm-1	Functional groups	Name of the Functional groups
3356	O-H	Alcohol
2924	C-H	Aliphatic
2852	C-H	Aliphatic
1637	C=O	Flavones
1099/1029	C-O stretching	Alcohols/ Phenols
779	=C-H bending	(out-of-plane bending) cis -RCH=CHR

Figure no 4: FTIR spectrum of *Sesuviumportulacastrum* leaf(SeSL)

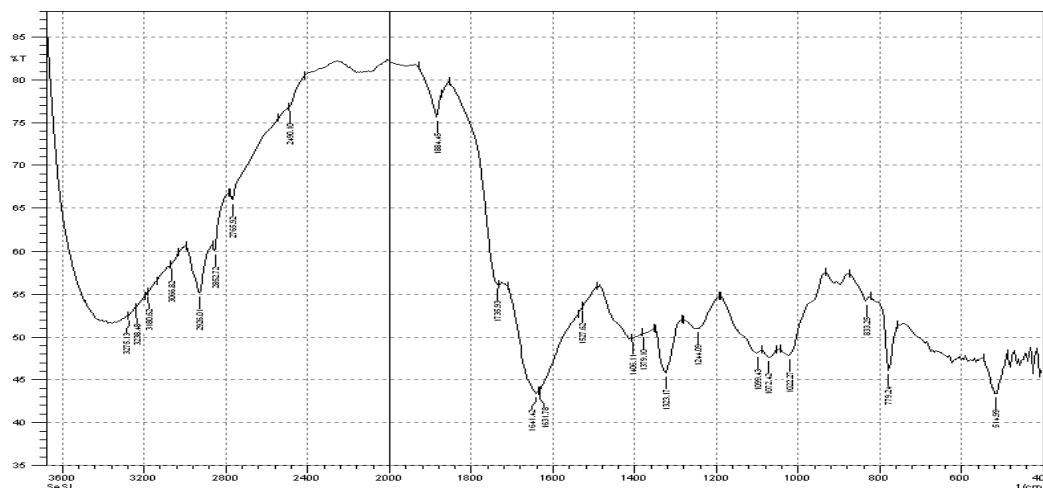


Figure no 5: FTIR spectrum of *Sesuviumportulacastrum* whole plant (S1)

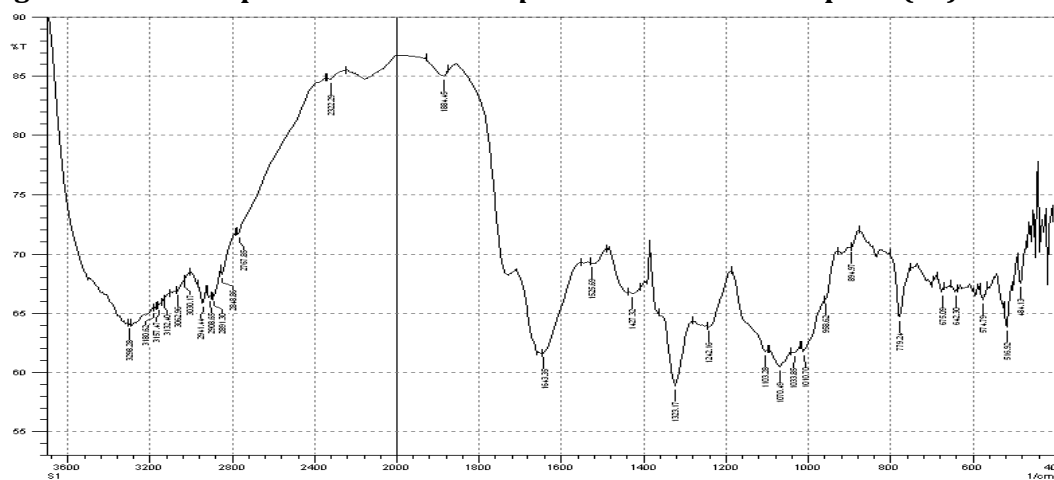
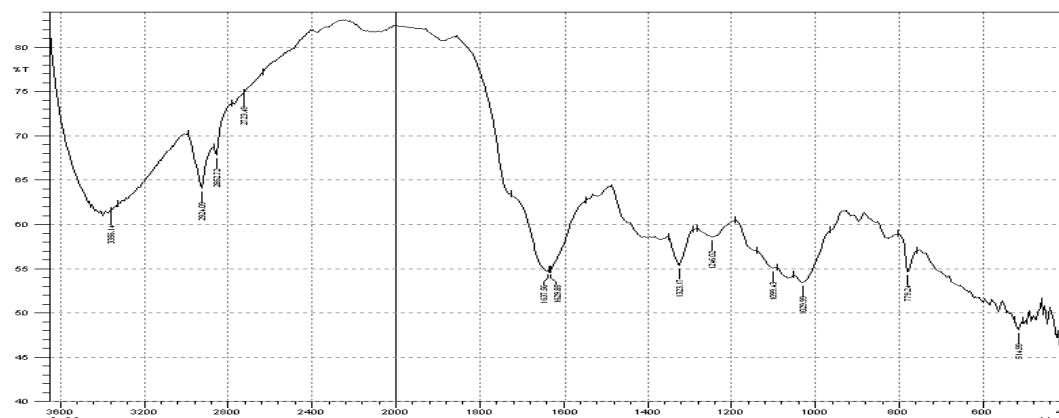


Table no 5: FTIR Result for *Sesuviumportulacastrum* whole plant (S1)

Wavelength in cm-1	Functional groups	Name of the Functional groups
3600-2400	O-H	Alcohol (very broad)
2941	C-H	Aliphatic
1641	C=C	Conjugated carbonyl (may be flavone)
1070	C-O	Alcohols/ Phenols
779	=C-H bending	(out-of-plane bending)cis -RCH=CHR

Figure no 6: FTIR spectrum of *Sesuviumportulacastrum* stem (SeSS)**Table no 6: FTIR Result for *Sesuviumportulacastrum* stem (SeSS)**

Wavelength in cm-1	Functional groups	Name of the Functional groups
3356	O-H	Alcohol
2924 .2852	C-H	Aliphatic
1629/1637	C=C	Arenes (In flavones C=O ,1637)
1099/1029	C-O stretch	Alcohols/ Phenols
898,723	=C-H bending	(out-of-plane bending) cis -RCH=CHR

References:

1. K.T Augusti,1996, Therapeutic values of onion and garlic. Indian. J. Exp .Biol., 34, pp 634-640.
2. Kathiresan K., Qasim S.Z. (2005). Biodiversity of Mangrove Ecosystems,1st Edn., Hindustan Publishing Corporation (India), New Delhi. 251.
3. Harbone JB. (1984). Phytochemical methods. Chapman and Hall. 2nd edition.
4. Pimpliskar MR, Jadhav RN, Jadhav BL,2012, Evaluation of Antimicrobial principles of Rhizophora species along Mumbai Coast, Journal of Advanced Scientific Research, 3,pp30-33.
5. ShariefMd N, Rao VU, 2011, Antibacterial activity of stem and root extracts of *Avicenniaofficinalis*L ,International Journal of Pharmaceutical Applications ,4, pp 231-236.
6. Asha KK, Mathew S, Lakshman PT,2012, Flavanoids and phenolic compounds in two mangrove species and their antioxidant property, Indian journal of geo-marine sciences, 41,pp259-264.
7. Lokhande VH, 2009a,*Sesuviumportulacastrum* (L.) L. a promising halophyte: cultivation, utilization and distribution in India, Genet Resour Crop Evol , 56,pp 741-747.
8. Lokhande VH, Nikam TD, Patade VY, Suprasanna P ,2009b, Morphological and molecular diversity analysis among the Indian clonesof*Sesuviumportulacastrum*L. Genet Res Crop Evol ,56, pp 705-717.
9. Magawa ML, Gundidza M, Gweru N, Humphrey G,2006, Chemical composition and biological activities of essential oil from the leaves of *Sesuviumportulacastrum*, J Ethnopharmacol ,103 pp 85- 89.
10. Padmakumar K, Ayyakkannu K ,1997, Seasonal variation of antibacterial and antifungal activities of the extracts of marine algae from Southern coast of India. Bot Mar, 40,pp 507-515.

11. Ghnaya T, Slama I, Messedi D, Grignon C, Ghorbel MH, Abdelly C, 2007, Cd-induced growth reduction in the halophyte *Sesuvium portulacastrum* is significantly improved by NaCl. *J Plant Res*, 120, pp 309–316.
12. Snehal B. Gagare, Pratima S. Jadhav, 2017, Molecular phylogeny of *Sesuvium portulacastrum* using 18S nuclear ribosomal gene sequence, *Global journal for research analysis*, 6 (5).
13. D.E. Okwu, 2004 Phytochemicals and Vitamin content of Indigenous Spices of South Eastern Nigeria, *J. Sustain. Agric. Environ*, 6(3), pp 34.
14. Pronob Gogoi and M. Islam. Phytochemical Screening of *Solanum nigrum* L and *S. myriacanthus* Dunal from Districts of Upper Assam, India. *IOSR Journal of Pharmacy*, 2012, 2, pp 455-459
15. Osborne, Brian G. (2006). Near-Infrared Spectroscopy in Food Analysis. John Wiley & Son
16. Mohamed Yacoob Syed Ali, Venkatraman Anuradha, Syed Abudhair Sirajudeen, Prathasarathy Vijaya, Nagarajan Yogananth, Ramachandran Rajan, Peer Mohamed Kalitha Parveen, 2013, Mosquito larvicidal properties of volatile oil from salt marsh mangrove plant of *Sesuvium portulacastrum* against *Anopheles stephensi* and *Aedes aegypti*, *Journal of Coastal Life Medicine*; 1(1), pp 66-71
17. Amad Al Azzawi, Alva Alqboori, Mahmood Hachim, 2012, Preliminary phytochemical and antibacterial screening of *Sesuvium portulacastrum* in United Arab Emirates, *Pharmacognosy Res*, 4(4), pp 219-224.
18. Harwood L M., Moody C. J., (1989). *Experimental organic chemistry: Principles and Practice (Illustrated ed.)*. Wiley-Blackwell, 292.
19. J.B. Harborne, (1973). *Phytochemical Methods*, (London: Chapman and Hall Ltd), 49-188.
20. G.E. Trease and W.C. Evans, *A textbook of Pharmacognosy*, 13 (London: Baillière Tindall Ltd), 1989, 19-21.
21. Chinnappan Ravinder Singha, Natarajan Sithranga Boopathya, Kandasamy Kathiresana, Sekar Anandhana, Sunil Kumar Saha, Aswin Kumar Ba, 2013, Effect of Bioactive Substances from Mangroves on Anti-Oxidant, Anti-Bacterial Activity and Molecular Docking Study Against Lung and Oral Cancer, *Photon*, 139, pp 226-236.
22. A. D. Bholay, Mayur Ingale, Apurv Gaur, 2015, Therapeutic potential of mangrove and its associate plant extracts from Thane Creek, against human respiratory tract MDR pathogens, *JBES*, 7(4), pp 118-126.
23. Trivedi MK, Panda P, Sethi, KK, Jana S., 2016, Metabolite profiling of *Withania somnifera* roots hydroalcoholic extract using LC-MS, GC-MS and NMR spectroscopy, *Chem Biodivers*, Article in press. doi:10.1002/cbdv.201600280
24. Tokusoglu O., Unal M.K., Yildirim Z., 2013, HPLC-UV and GC-MS characterization of the flavonol glycosides Quercetin, Kaempferol and Myricetin in Tomato pastes and other tomato-based products, *Acta Chromatograph*, 13, pp 196-207.
25. Betz JM, Gay ML, Mossoba MM, Adams S and Portz BS, 1997, Chiral gas chromatographic determination of ephedrine-type alkaloids in dietary supplements containing MáHuáng, *J AOAC Int*, 80, pp 303 -15.